Chemical respiratory allergy is an important occupational health problem, but there are currently available no validated methods for hazard identification. This is due in part to the fact that the relevant cellular and molecular mechanisms of sensitization of the respiratory tract have been unclear, with particular controversy regarding the role of IgE. There is now increasing evidence that respiratory sensitization is associated with the preferential activation of type 2 T lymphocytes and the expression of type 2 cytokines interleukin (IL)-4, IL-5, IL-10, and IL-13. Type 2 cell products favor immediate type hypersensitivity reactions, serving as growth and differentiation factors for mast cells and eosinophils, the cellular effectors of the clinical manifestations of the allergic responses, and promoting IgE antibody production. There has been considerable interest in the application of cytokine profiling for the characterization of chemical allergens, with cytokine phenotypes analyzed in freshly isolated tissue, or following culture in the presence or absence of mitogen at the level of protein secretion or mRNA expression. Experience to date suggests that the measurement of induced cytokine secretion profiles shows promise for the hazard identification and characterization of chemical respiratory allergens. The purpose of this brief review article is to consider the approaches available and to highlight key procedural issues.

Key Words: chemical respiratory allergy; hazard identification; IgE; type 2 cell products; cytokine profiling.
atic patients displaying measurable IgE (Tarlo, 1999; Tee et al., 1998). Despite uncertainty regarding a universal association between IgE antibody and occupational chemical respiratory allergy, there is increasing evidence that allergic sensitization of the respiratory tract will be favored by the induction of a polarized type 2 immune response (Kimber and Dearman, 1997; Mapp et al., 1999). Type 2 cells produce cytokines that facilitate the development and expression of immediate type hypersensitivity reactions, including respiratory sensitization. In common with the development of asthma to protein allergens (Durham et al., 1992; Robinson et al., 1993), eosinophils and T lymphocytes are recruited into the site of inflammation (Bentley et al., 1992) and there is evidence that such lymphocytes selectively express type 2 cytokines (Del Prete et al., 1993; Maestrelli et al., 1994).

With the increased understanding of the immunobiology of chemical respiratory allergy have come new opportunities for the development of tests for the identification of potential respiratory allergens: tests that do not rely upon the elicitation of clinical manifestations of respiratory distress. The aim of this article is to consider the practical application of one such test, cytokine profiling, for the characterization and identification of chemical allergens.

**Immunotoxicology of Allergic Responses**

Before considering the practical application of cytokine fingerprinting, it is necessary to review briefly the immunological mechanisms that the method attempts to model. Low molecular weight chemical allergens in their native state are unable to induce immune responses and in order to be recognized as foreign by the immune system and to initiate an immune response, a protein-chemical complex must be formed (Landsteiner and Jacobs, 1936). These complexes are internalized and processed for presentation to other cells of the immune system by antigen-presenting cells, with dendritic cells being the most efficient and the only cells that can initiate primary immune responses (Viola et al., 1999). In response to antigen, specific B lymphocytes are stimulated to divide and differentiate into plasma cells that have the machinery to synthesize and secrete specific antibody, including IgE antibody. However, the development of immune responses, including allergic responses, is orchestrated by the activity of CD4+ T helper (Th) cell subpopulations and their cytokine products. In the mature immune response, two phenotypes of Th cell, designated Th1 and Th2, predominate and are distinguished as a function of the cytokines they secrete (Abbas et al., 1996; O’Garra, 1998; Romagnani, 2000). While both populations secrete interleukin (IL) 3 and granulocyte/macrophage-colony stimulating factor (GM-CSF), only Th1 cells express interferon γ (IFN-γ), IL-2, and tumor necrosis factor-β (TNF-β), and only Th2 cells express IL-4, IL-5, IL-6, IL-10, and IL-13. The existence of functional subpopulations of Th cells provides a mechanism for the development of different qualities of immune responses depending upon the nature of the challenge to the host. Th1 cells are adapted for the provision of cell-mediated immunity, whereas Th2 cells produce cytokines that promote humoral immune function and the costimulation and differentiation of B lymphocytes (Abbas et al., 1996; Mosmann and Sad, 1996). This functional dichotomy of Th cells is relevant for the development and expression of allergic disease. Th1 cells are associated with DTH responses, with adoptive transfer of Th1 cell clones sufficient to transfer DTH reactions to naïve recipients (Cher and Mosmann, 1987) and the development of such responses being facilitated by the Th1 cytokine IFN-γ (Diamantstein et al., 1988). Conversely, Th2 cell activation promotes immediate type hypersensitivity reactions, with expression of the type 2 cytokine IL-4 essential for the induction and maintenance of IgE antibody responses (Finkelman et al., 1988b; Kuhn et al., 1991). In contrast, the type 1 cell product IFN-γ inhibits IgE production (Finkelman et al., 1988a). In addition, type 2 cytokines promote other aspects of the immediate type reactions, serving as growth and differentiation factors for mast cells and eosinophils and stimulating the recruitment of eosinophils, cells that play important roles in the elicitation of the clinical manifestations of the allergic response (Kimber and Dearman, 1997; Krishnan and Mosmann, 1998). These polarized forms of Th cell develop with time or following repeated exposure from precursor cells, designated Th0 cells, which have the potential to express both Th1 and Th2 type cytokines, and which themselves develop from primitive precursor (Thp) cells that express IL-2 only (Bendelac and Schwartz, 1991; Mosmann et al., 1991). The differentiation of Th1 and Th2 cells is controlled largely by positive feedback and negative cross-regulation by Th1 and Th2 cell products (Krishnan and Mosmann, 1998). The cytokines that appear to be of greatest significance in polarizing the immune response are the type 2 cell product IL-10, and IL-12, a heterodimeric cytokine produced by dendritic cells and some epithelial cells. The source of the cytokines that drive the initial development of divergent Th cell phenotypes is still a matter for some debate, but it has been suggested that the cellular components of the innate immune system, including mast cells, may play a role (Kimber and Dearman, 1997). Therefore it is the balance between Th1 and Th2 cytokines that determines the direction and nature of the induced immune response.

Although attention has focused on the role of Th cells and their products in the development of allergic responses, it has become apparent that CD8+ T cytotoxic (Tc) cells may also play influential roles. Two populations, designated Tc1 and Tc2, have been described that display selective cytokine secretion patterns analogous with Th1 and Th2 cells, respectively (Mosmann and Sad, 1996). Although there is limited evidence of a physiological role for Tc2-type cells (Kimber and Dearman, 1997), there is increasing interest in the possibility that Tc1-type cells that produce IFN-γ may mediate and/or regulate contact hypersensitivity reactions (Kimber and Dearman, 2002). Until recently, allergic contact dermatitis, in common
with other forms of DTH reaction, was viewed as a Th1-type response, mediated by CD4^+ IFN-γ-producing effector cells. However, there is a growing appreciation that, in contrast to DTH reactions to complex protein or cellular antigens, CD8^+ effector cells may play important roles in the development and elicitation of allergic contact dermatitis to chemical allergens (Gorbachev and Fairchild, 2001). Allergic contact dermatitis to chemicals is apparently associated with the preferential activation of Th1 and Tc1 cells, whereas immediate type hypersensitivity reactions such as chemical respiratory hypersensitivity are favorably by the selective activation of Th2 (and possibly Tc2) type cells.

**Cytokine Profiling of Chemical Allergens: Background to Method Development**

The observation that topical exposure of BALB/c strain mice to chemical contact and respiratory allergens stimulates different qualities of immune response with respect to IgE and IgG antibody responses (Dearman and Kimber, 1991) led us to question whether it might be possible to characterize allergens as a function of cytokine secretion profiles. Under conditions of exposure of equivalent immunogenicity with respect to lymphocyte proliferation and IgG antibody responses, the cytokine phenotypes induced by the contact allergens 2,4-dinitrochlorobenzene (DNCB) and trimellitic anhydride (TMA) were examined (Dearman et al., 1997). Draining lymph node cells (LNC) excised from DNCB-treated mice expressed high levels of the type 1 cytokine IFN-γ, but relatively low levels of the type 2 cytokines IL-4 and IL-10. The converse type 2 cytokine secretion profile was provoked by topical exposure to TMA (Dearman et al., 1994). Interestingly, these polarized cytokine phenotypes take time to mature, with LNC isolated three days after initiation of exposure displaying a mixed Th0-like phenotype with type 1 and type 2 cytokines expressed following treatment with both chemical contact and respiratory allergens (Betts et al., 2002; Dearman et al., 1994; Moussavi et al., 1998). A more prolonged (13-day) exposure protocol was required for the development of a differentiated cytokine phenotype (Betts et al., 2002; Dearman et al., 1994, 1997). The relative contribution of CD4^+ and CD8^+ cells to the cytokine phenotype of allergen-activated cells has been examined using negative (complement depletion) and positive selection (antibody coated magnetic beads) techniques (Dearman et al., 1996; Moussavi et al., 1998). These experiments demonstrated that early (three days) following initiation of exposure to either DNCB or TMA, the mixed cytokine phenotype observed was a function of IFN-γ-secreting CD4^+ (Th1) and CD8^+ (Tc1) cells and IL-4-secreting CD4^+ (Th2) cells (Moussavi et al., 1998). The preferential type 2 cytokine secretion profile observed after prolonged treatment with the respiratory sensitizer TMA was due to the activation of CD4^+ (Th2) cells, with the relatively low levels of IFN-γ derived exclusively from CD8^+ (Tc1) cells. The selective type 1 cytokine expression pattern stimulated by more chronic exposure to DNCB was associated with both CD4^+ (Th1) and CD8^+ (Tc1) IFN-γ-expressing cells, with the low levels of type 2 cytokines being a result of CD4^+ (Th2) cell activation (Dearman et al., 1996). These data suggest that as the immune response to a chemical allergen matures with time and becomes polarized, exposure to the contact allergen DNCB is associated with the selective development of Th1 and Tc1 cells, whereas treatment with the respiratory allergen TMA is associated primarily with Th2 cell development.

**Initial Evaluation of Cytokine Profiling**

Subsequent investigations using a range of additional chemical respiratory allergens, including isocyanates, platinum salts, glutaraldehyde, and various acid anhydrides, have confirmed that the expression of a preferential type 2 cytokine expression profile is a general property of respiratory sensitizing chemicals (Dearman and Kimber, 2001). Using the same exposure protocol as that used for the chemical respiratory allergens described above, treatment with allergens such as 2,4-dinitrofluorobenzene (DNFB), isoeugenol, and hexyl cinnamic aldehyde (contact allergens that apparently lack respiratory sensitizing activity) stimulates instead a selective type 1 cytokine secretion pattern (Dearman and Kimber, 2001). For selected respiratory allergens, including the acid anhydrides phthalic anhydride (PA) and maleic anhydride, and the isocyanates diphenyl methane diisocyanate (MDI) and fluorescein isocyanate, it has been confirmed that the induced preferential type 2 cytokine expression profile is a function of Th2 (CD4^+ ) cell activation exclusively (Dearman and Kimber, 2000; Dearman et al., 1996, 2002a).

**Cytokine Profiling in Practice: Considerations of Dose Selection and Controls**

These data demonstrate that for a variety of chemical allergens, the measurement of induced secretion of IFN-γ and IL-12 provide preferential markers of type 1 responses, whereas IL-4, IL-5, IL-10, and IL-13 can be used as markers of selective type 2 responses. In the experiments described above, LNC have been pooled on an experimental group basis and single cell suspensions prepared for culture. This is in order to provide for a sufficient yield of LNC for the analysis of the complete range of cytokines that would not be possible if LNC were prepared on an individual animal basis. For the reference contact allergen DNCB and respiratory allergen TMA, despite some interexperimental variations in absolute levels of cytokine, analysis of cytokine phenotypes revealed that the ability of DNCB and TMA to induce Th1 and Th2 cytokine profiles, respectively, was very consistent. Statistical analyses (Student’s t-test) of four independent experiments demonstrated that differences in cytokine expression patterns between DNCB and TMA-activated LNC were significant at the 5% (IFN-γ, IL-12, and IL-10) or 1% (IL-4, IL-5, and IL-13) levels.
LLNA are very unlikely to have a significant potential to cause respiratory sensitization if similar levels of type 2 cytokines are not inconsistent with the results reported herein, as spontaneous expression of type 2 cytokines was also detected after treatment with DNCB, albeit at much lower levels than those stimulated by TMA (Dearman et al., 1997; Hayashi et al., 2001). However, it is not possible in the experiments conducted by Ulrich et al. (2001) to compare directly the ability of DNCB and TMA to provoke selective type 1 and type 2 cytokine secretion patterns since the concentration of DNCB used was considerably less immunogenic than was the dose of TMA selected.

Considerations of Strain and Species

In the majority of the experiments described above, BALB/c mice have been used (Dearman and Kimber, 2001; Hayashi et al., 2001; Manetz et al., 2001; Ulrich et al., 2001; Vandebriel et al., 2000; van Och et al., 2002). Mice of this strain are known to be high IgE responders and are predisposed to make Th2-type responses. It could be argued that the use of the BALB/c strain mouse may have biased the observed cytokine phenotype towards type 2 responses. It must be remembered that concurrent exposure of this mouse strain to contact allergens such as DNCB under conditions of equivalent immunogenicity provokes selective type 1 cytokine responses (Dearman and Kimber, 2001; Hayashi et al., 2001; Manetz et al., 2001; Vandebriel et al., 2000; van Och et al., 2002). Furthermore, recent investigations have demonstrated that in a mouse strain that is more predisposed to make type 1 responses, similar divergent patterns of cytokine expression are elicited by DNCB and TMA (Hayashi et al., 2001). C57BL/6 strain mice are reported to be biased towards type 1 responses (Guler et al., 1996), however, exposure of this strain of mice to respiratory allergens (including TMA, PA, and MDI) also stimulated a preferential type 2 cytokine response, whereas contact allergens such as DNCB and oxazolone induced a selective type 1 cytokine phenotype (Hayashi et al., 2001). In some experiments, an alternative rodent strain has been employed, the Brown Norway (BN) rat, which in common with the BALB/c strain mouse is regarded as having a predisposition to type 2 responses. Topical exposure of BN rats to DNCB or to TMA under conditions of equivalent immunogenicity with respect to induced proliferative responses resulted in significant increases in total serum IgE concentration being recorded only following treatment with TMA (Arts et al., 1997). In addition, measurement of cytokine secretion profiles provoked in draining LNC...
by topical exposure of BN rats to DNCB or to TMA revealed the selective induction of type 1 and type 2 cytokine patterns, respectively (Dearman et al., 2002b). Taken together, these data suggest that chemical contact allergens such as DNCB and respiratory sensitizers such as TMA exhibit an innate ability to stimulate type 1 and type 2 cytokine production profiles, respectively, which are species and strain independent.

**Route of Exposure: Is the Skin Appropriate?**

Another important issue is that of route of exposure. In the experiments described above, rodents have been treated with chemical allergen epicutaneously and there are reports in the literature that prolonged topical exposure and repeated applications of chemical allergen may result in a bias towards Th2-type responses (Kitagaki et al., 1999). However, the exposure regimen utilized in the majority of the experiments described above has been designed to induce polarized cytokine secretion profiles following topical exposure to different classes of chemical allergen (Dearman and Kimber, 2001; Hayashi et al., 2001; Manetz et al., 2001). Thus, repeated topical application over a 13-day period of the known human contact allergen DNCB results in the development of a selective type 1 cytokine phenotype (Dearman and Kimber, 2001; Hayashi et al., 2001; Manetz et al., 2001). By definition, the skin represents the relevant route of exposure for contact sensitization; there is reason to believe, however, that dermal exposure may be relevant also for induction of effective sensitization of the respiratory tract. Although inhalation is undoubtedly the most important and most common route of exposure to chemical respiratory allergens, there is experimental evidence and some limited clinical evidence to suggest that sensitization of the respiratory tract may be induced following skin contact (Kimber and Dearman, 2002). Further it has been demonstrated that similar qualities of immune response with respect to antibody isotype profiles are stimulated in BALB/c strain mice regardless of whether sensitization is via inhalation or the skin, with only the respiratory allergen TMA inducing IgE antibody expression (Dearman and Kimber, 1991; Dearman et al., 1991). Taken together with the fact that the dermal route of exposure facilitates analyses of cytokine responses induced in discrete draining lymph nodes, our accumulated experience confirms that the characterization of cytokine responses induced following topical application of different classes of chemical allergen is an appropriate strategy for hazard identification.

**Requirement for Restimulation in Vitro?**

One of the protocol variations that has been explored by several investigators is to measure cytokine expression (either at the protein secretion or mRNA level) following restimulation of LNC with mitogen in vitro (Manetz et al., 2001; Ulrich et al., 2001; Vandebriel et al., 2000; van Och et al., 2002). Although it is necessary to restimulate LNC with the T cell mitogen concanavalin A (con A) in order to detect expression of IL-4 protein, there is sufficient spontaneous production of the other cytokines (IFN-γ, IL-5, IL-10, IL-12, and IL-13) for measurement by enzyme-linked immunosorbent assay (ELISA) in the absence of restimulation (Dearman et al., 1994, 2002a,b). Mitogen activation per se does not stimulate the expression of IL-4. Thus, cells derived from naïve or vehicle-treated mice fail to secrete measurable levels of IL-4, despite the presence of mitogen (Dearman et al., 2002b). Other authors have described similar difficulties in the detection of IL-4 and have suggested that such may be due to the uptake of this cytokine in autocrine fashion, given that measurement of IL-4 is more efficient when reutilization of this cytokine is blocked by antireceptor antibodies (Bancroft et al., 1998). There is some evidence to suggest that the addition of mitogens such as polyclonal anti-CD3 antibody may bias the cytokine secretion profile with respect to cytokines other than IL-4; certainly the experience of Ulrich et al. (2001) that contact allergens induce type 1 and type 2 cytokine expression may be explained partly by the incorporation of a mitogen restimulation step. Indeed, other investigators have shown that stimulation of naïve spleen cells with anti-CD3 antibody is sufficient to induce detectable cytokine production (Wang et al., 1992). In contrast, with the exception of IL-12 protein, which is produced constitutively, quiescent control LNC do not generally produce detectable levels of other cytokines (IFN-γ, IL-4, IL-5, IL-10, and IL-13) in the absence of mitogen restimulation, suggesting that the measurement of cytokine expression in the absence of mitogen may be more selective for allergen-induced effects (Betts et al., 2002; Dearman et al., 1994, 2002a). Under some circumstances restimulation with mitogen may not compromise completely the development of polarized cytokine phenotypes. Culture of LNC isolated five to seven days after initiation of exposure to different classes of chemical allergen with the T cell mitogen con A does result in the induction of divergent cytokine expression profiles (Vandebriel et al., 2000; van Och et al., 2002). However, such cytokine secretion patterns are less polarized than those derived following measurement of spontaneous cytokine production, particularly with respect to IFN-γ expression, suggesting that analysis of cytokine profiles in the absence of restimulation may provide a more robust assessment of sensitizing hazard.

**Message or Protein?**

One other area of significant variation among cytokine profiling protocols is whether cytokine responses are measured as a function of protein secretion, or at the level of mRNA expression. Cytokine mRNA expression has been examined by reverse transcription polymerase chain reaction (RT-PCR; Hayashi et al., 2001; Ryan et al., 1998; Vandebriel et al., 2000; Warbrick et al., 1998) or by ribonuclease protection assay (RPA; Betts et al., 2002; Manetz et al., 2001; Plitnik et al., 2002). Measurement of cytokine transcripts by RT-PCR re-
revealed that in the absence of mitogen restimulation in vitro, prolonged exposure to TMA induced increased levels of IL-4 mRNA expression compared with treatment with DNCB (Hayashi et al., 2001; Ryan et al., 1998; Warbrick et al., 1998). The reverse pattern of cytokine mRNA expression was recorded for the type 1 cytokines IFN-γ (Warbrick et al., 1998) and IL-12 (Hayashi et al., 2001), although it must be noted that in the latter studies and in other studies, IFN-γ mRNA levels did not discriminate between contact and respiratory allergens (Hayashi et al., 2001; Vandebriel et al., 2000). Several studies have now been published in which allergen-induced changes in cytokine gene expression have been analyzed by RPA, a technique with a somewhat lower level of sensitivity than RT-PCR (Betts et al., 2002; Manetz et al., 2001; Plitnik et al., 2002). This method is sufficiently sensitive to detect consistently preferential type 2 cytokine expression following prolonged treatment with the respiratory allergens TMA and toluene diisocyanate (TDI), with particularly high levels of transcripts for IL-4 and IL-10 detected in freshly isolated tissue (Betts et al., 2002; Manetz et al., 2001; Plitnik et al., 2002). Without restimulation in vitro with T cell mitogen con A, RNA isolated from DNCB- or DNFB-activated LNC tissue did not express a selective type 1 cytokine mRNA phenotype, particularly with respect to IFN-γ (Manetz et al., 2001; Plitnik et al., 2002). The reasons for the failure to detect increased mRNA for IFN-γ despite robust secretion of this cytokine are presently unclear, although it would appear that production of this cytokine by draining LNC is controlled mainly at the level of secretion. The measurement of cytokine mRNA levels in freshly isolated tissue by RPA offers a number of potential advantages compared with other methods for cytokine analysis, including the lack of requirement for tissue culture or restimulation, the simultaneous measurement of multiple cytokines and the possibility of evaluating cytokine responses at the individual animal level, given the relatively small amounts of tissue used. It should be noted that to date analysis of cytokine expression by RPA has only been performed on tissue derived following treatment with a very limited number of chemical allergens (Betts et al., 2002; Manetz et al., 2001; Plitnik et al., 2002). These experiments revealed that whereas the two respiratory allergens tested could be identified on the basis of increased levels of mRNA for type 2 cytokines in freshly isolated tissue, there was no measurable induction by contact allergens of transcripts for type 1 cytokines in the absence of restimulation (Betts et al., 2002; Manetz et al., 2001; Plitnik et al., 2002).

With respect to the characterization of sensitizing hazard, the judicious view is that cytokine profiling by RPA and by RT-PCR may identify those chemicals with respiratory sensitizing potential as a function of induced type 2 cytokine expression. More experience is required with a wider range of respiratory sensitizers before the sensitivity and selectivity of this method versus measurement of protein secretion by ELISA can be assessed. However, as both methods are less effective at measuring the expression of type 1 cytokines such as IFN-γ than is measurement of protein secretion by ELISA, they have less utility for the identification of contact allergens and the discrimination between respiratory and contact allergens.

**Interexperimental Variation**

Another important issue is that of interexperimental variation. In the most commonly applied configuration of cytokine profiling (Betts et al., 2002; Dearman and Kimber, 2001; Dearman et al., 2002a), as described previously, LNC are pooled on an experimental group basis and cultured for preparation of supernatants and analysis of cytokine protein secretion by ELISA. It is not possible therefore to examine interanimal variations in cytokine production patterns. However, as described in a previous section (Cytokine Profiling in Practice: Considerations of Dose Selection and Controls), the statistical significance of interexperimental differences in cytokine secretion phenotypes of DNCB- and TMA-activated LNC has been addressed. These studies demonstrated that DNCB-stimulated LNC expressed significantly higher levels of the Th1-type cytokines IFN-γ and IL-12 under conditions where TMA-activated LNC produced significantly higher levels of the Th2-type cytokines IL-4, IL-5, IL-10, and IL-13 (Dearman and Kimber, 2001; Dearman et al., 2002b). Using an alternative method in which LNC are restimulated with mitogen prior to analysis of cytokine protein production by ELISA, lymph nodes were pooled on an individual animal basis following exposure to allergen, allowing assessment of interanimal variation in cytokine expression, although insufficient cells were available from vehicle-treated lymph nodes to perform these analyses (Vandebriel et al., 2000). However, the addition of mitogen, as described previously, results in cytokine secretion profiles for DNCB- and TMA-activated cells that are less polarized than those observed following culture of LNC in the absence of mitogen (Dearman and Kimber, 2001; Vandebriel et al., 2000). The addition of mitogen, notwithstanding the advantage of provision for analysis of cytokine expression at an individual animal level, results in a considerable loss of sensitivity.

Recent advances in methodology for the measurement of cytokine proteins have made it possible to analyze the expression of multiple cytokines in small sample volumes, facilitating the measurement of cytokine secretion patterns at the individual animal level in the absence of restimulation. Systems such as the Bio-Plex™ cytokine assay (Bio-Rad, Hercules, CA) are based on a similar principle to a capture sandwich ELISA, but anticytokine antibodies are covalently coupled to dyed polystyrene beads in a microtiter format. Relatively small volumes of sample (50 µl) are required and bound cytokine is visualized using biotinylated detection antibody and phycoerythrin-la beled streptavidin. By reading each individual bead in the mixture, the analytical system can discriminate between up to 100 different proteins including cytokines. Figure 1 displays the cytokine secretion profiles for LNC derived from DNCB-
and TMA-exposed mice (n = 5), where lymph nodes have been pooled on an individual animal basis and cultured for 120 h in the absence of mitogen. Production of the type 1 cytokines IFN-γ and IL-12 (p40 and p70) and IFN-γ expression than did the other DNCB-treated animals. Even so, between group comparisons revealed that LNC derived from DNCB-exposed animals produced significantly more IFN-γ (p < 0.05) and IL-12 p40 (p < 0.01) than did those isolated from TMA-treated mice, whereas exposure to TMA stimulated significantly higher levels of IL-4, IL-5, and IL-10 (p < 0.01). Interestingly, this system was also sensitive enough to detect IL-4 secretion in the absence of restimulation with mitogen. These data suggest that there is little interanimal variation in the ability of DNCB and TMA to elicit distinct type 1 and type 2 cytokine secretion profiles and demonstrate that similar cytokine production patterns are detected regardless of whether lymph nodes are pooled on a group basis or individual animals are assessed.

Concluding Comments

Substantial progress has been made with respect to the understanding of the immunotoxicology of chemical respiratory allergy. There is an increasing consensus that this important occupational disease is a result of the induction and activation of preferential type 2 T lymphocyte responses following exposure to chemicals with respiratory sensitizing potential. There has been interest in the application of cytokine profiling to the identification and characterization of chemical allergy, with cytokine phenotypes analyzed in freshly isolated tissue or after culture in the presence or absence of mitogen at the level of protein secretion or mRNA expression. The most common configuration of cytokine fingerprinting (measurement of induced cytokine secretion patterns in the absence of restimulation) in particular shows considerable promise as an approach for the identification of chemical respiratory allergens.

REFERENCES


Chan-Yeung, M., and Malo, J.-L. (1993). Compendium 1. Table of major occupational disease is a result of the induction and activation of preferential type 2 T lymphocyte responses following exposure to chemicals with respiratory sensitizing potential. There has been interest in the application of cytokine profiling to the identification and characterization of chemical allergy, with cytokine phenotypes analyzed in freshly isolated tissue or after culture in the presence or absence of mitogen at the level of protein secretion or mRNA expression. The most common configuration of cytokine fingerprinting (measurement of induced cytokine secretion patterns in the absence of restimulation) in particular shows considerable promise as an approach for the identification of chemical respiratory allergens.


NIH (1999). The murine local lymph node assay: A test method for assessing the allergic contact dermatitis potential of chemicals/compounds. NIH Pub-


