

Development and evaluation of PCR-based diagnostics for identification of *Salmonella* O antigens based on the *rfb* locus

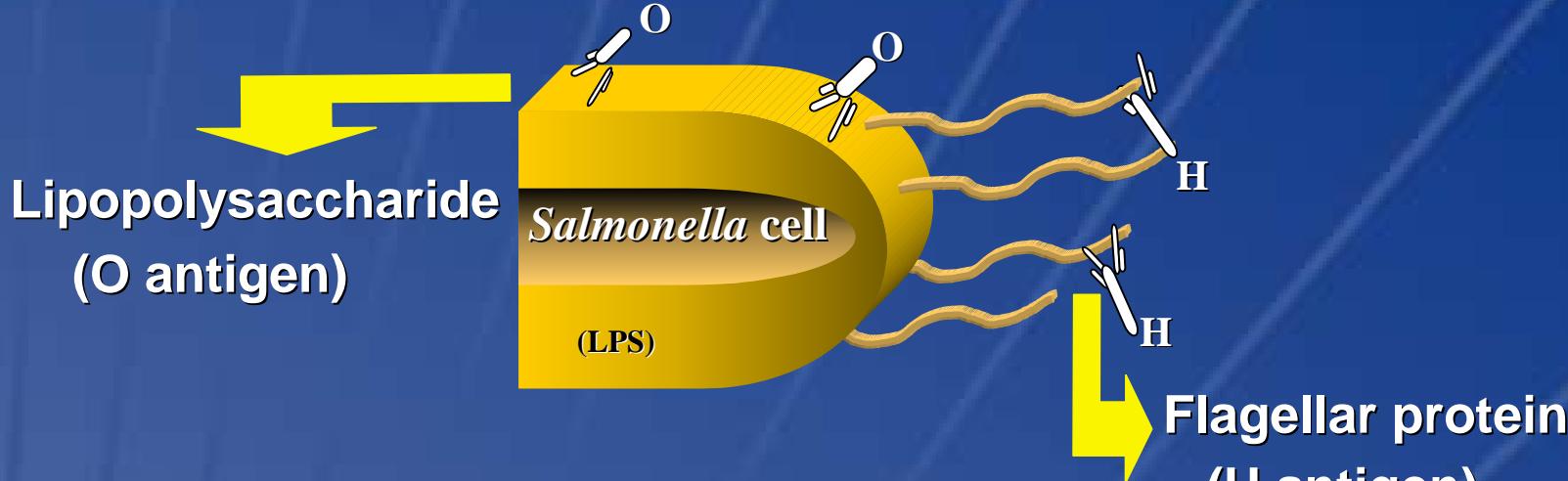


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What is serotyping ?

- Subtyping based on antigenic reactivity of:



- Designated by names or formulas
(subspecies O: Phase1 H : Phase 2 H)

S. Typhimurium: I 1,4,[5],12: i:1,2

O- antigens H-antigens

2501 serotypes

Why is it important ?

- Serotyping is very useful for the epidemiologic classification of strains
- It is the basis of the National *Salmonella* Surveillance System

Technical issues

- Reagents required are expensive and time consuming to produce and quality control
 - 250 O and H grouping, typing and single factor reagents needed to identify all serotypes
 - 350 stock strains and antigens used to evaluate and quality control antisera

Main Objective

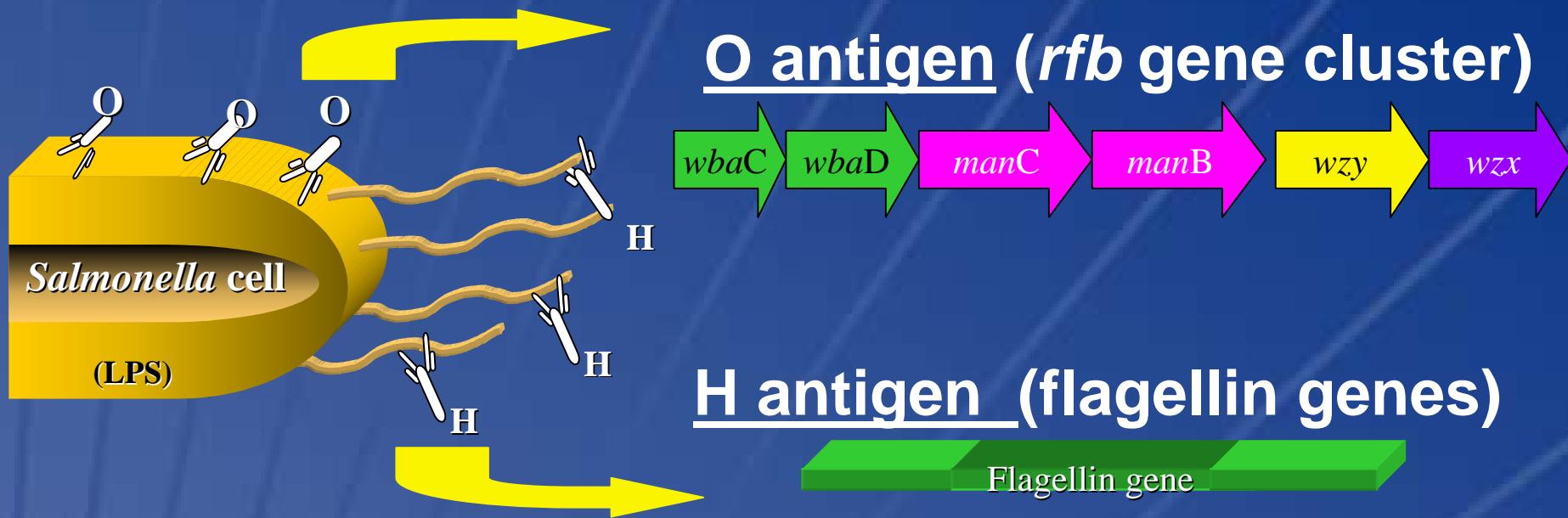
- Develop a DNA-based system that can replace traditional methods

“Molecular Serotyping”

- The new system should correlate with the current serotyping scheme so that results from the two systems can be compared

Molecular serotyping goals

- Characterize and/or sequence genes required for biosynthesis of:

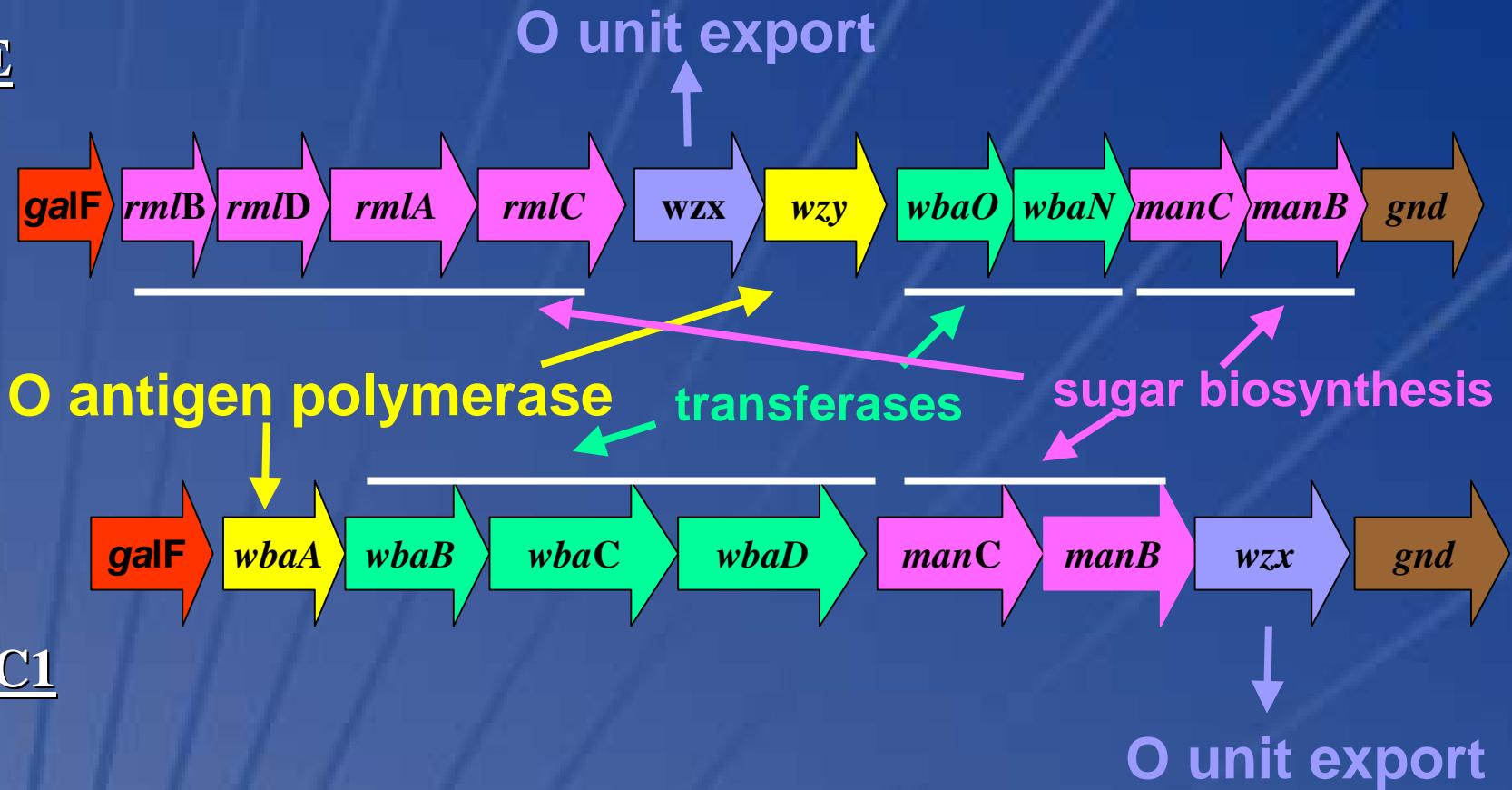


- Identify antigen-specific sequences
- Develop the target sequence into a molecular probe using appropriate technology

Salmonella O antigen gene clusters

- O antigens are encoded primarily by the *rfb* region

E



C1

Background

- ~~Identify and cluster~~- Selective amplification of sugar biosynthesis genes (*rha*, *prt*) to identify serogroups has been partially/totally sequenced for 9 serogroups A/D ($A_1, B_1, C_1, C_2, D_1, D_2, D_3, E_1$ and O:54)
 - no specific PCR has yet been described for serogroups C_1 , E or O:54

AIMS

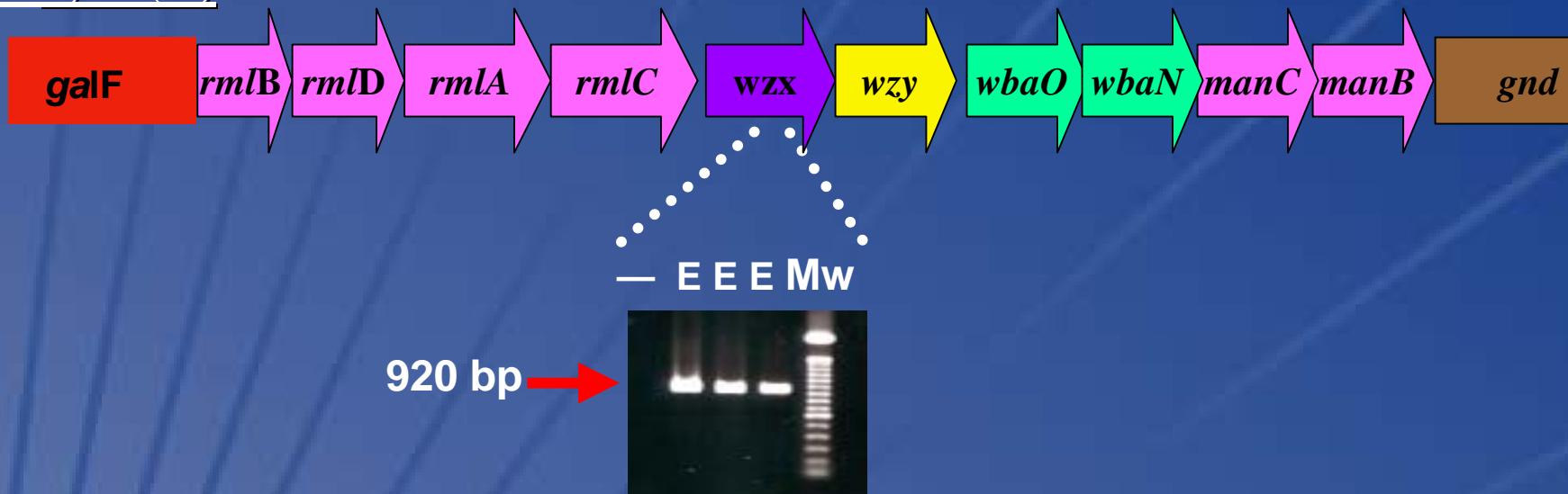
- Develop serogroup specific PCR's for those *rfb* gene clusters already sequenced
- Characterization of *rfb* gene clusters from additional serogroups

Serogroup identification by PCR (1)

O:6,7 (C1)



O:3,10 (E)



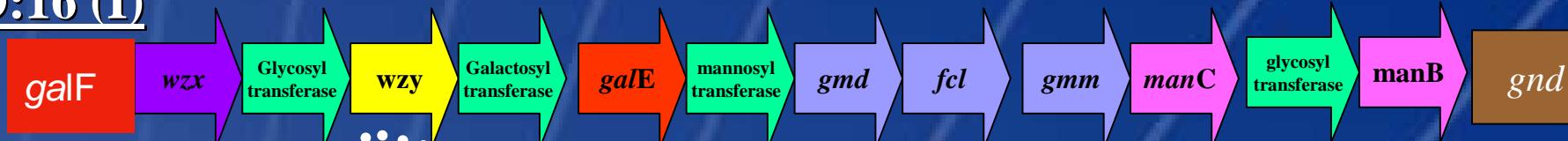
DNA sequence analysis of *rfb* gene clusters

Serogroup	Serotype	<i>rfb</i> size
O:6,14 (H)	<i>S. Sundsvall</i>	8.436 kb
O:16 (I)	<i>S. Gaminara</i>	12.891 kb
O:17 (J)	<i>S. Jangwani</i>	9.195 kb
O:18 (K)	<i>S. Cerro</i>	9.135 kb
O:35 (O)	<i>S. Ealing</i>	13.642 kb
O:38 (P)	<i>S. Roan</i>	6.013 kb
O:41 (S)	<i>S. Ipswich</i>	9.578 kb
O:48 (Y)	<i>S. Marina</i>	13.079 kb

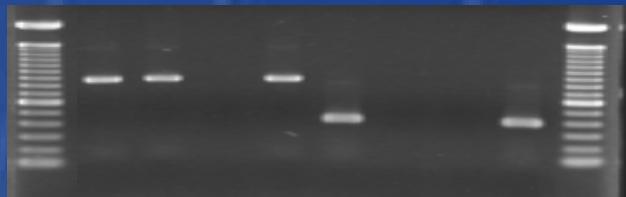
Characterization of *rfb* gene clusters from additional serogroups for which we currently have no sequence information

Serogroup identification by PCR (2)

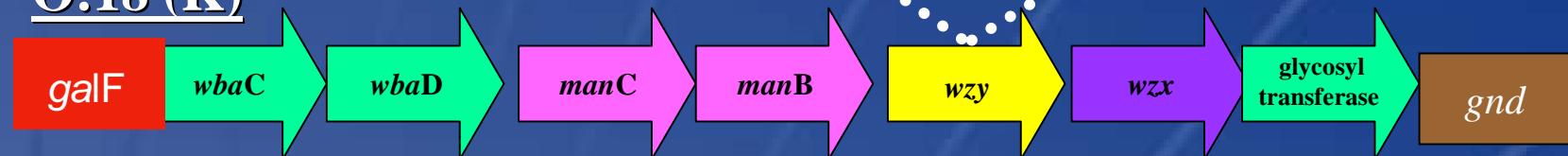
O:16 (I)



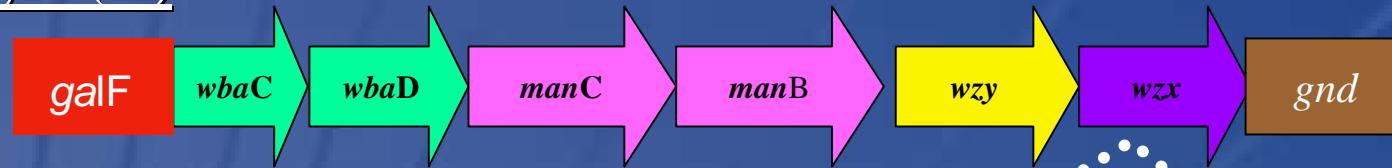
Mw I I - I K D₁ G K Mw



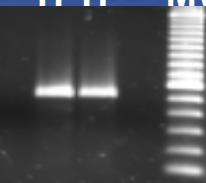
O:18 (K)



O:6,14 (H)



501 bp →



Serogroup	Serotype	Target	Primers evaluated against 46 serogroups (398 strains)
C ₁ (6,7,14)	S. Montevideo	wbaA	y
E (3,10,15)	S. Anatum	wzx	y
O:54	S. Borreze	wbbF	y
H (0:6,7,24,25)	S. Sundsvall	wzx	y
I (O:16)	S. Gaminara	wzy	y
J (O:17)	S. Jangwani	wzy	y
K (O:18)	S. Cerro	wzy	y
O (O:35)	S. Ealing	wzx	y
P (O:38)	S. Roan	wzy	y
S (O:41)	S. Ipswich	wzx	y
Y (O:48)	S. Marina	wzx	y
A/D	S. Paratyphi A / S. Typhi	prt	y
B	S. Typhimurium	abe	y
C ₂	S. Muenchen	abe	y

Phase I objectives

- Molecular identification of top 100 serotypes
 - 98% of clinical isolates

O:11 (F) *S. Abaetetuba*

O:30 (N) *S. Urbana*

O:13 (G) *S. Poona*

O:40 (R) *S. Johannesburg*

O:21 (L) *S. Minnesota*

O:43 (U) *S. Houten*

O:28_{ab} (M) *S. Pomona*

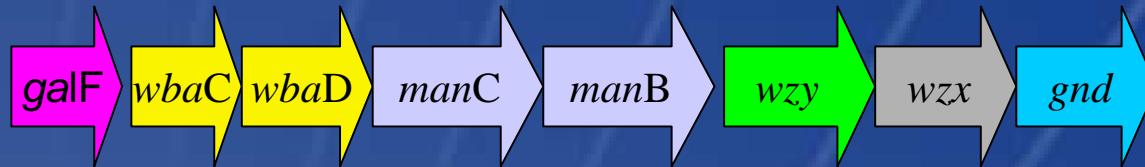
O:47 (V) *S. Bere*

O:28_{ac} (M) *S. Dakar*

O:50 (Z) *S. Flint*

Phase I objectives

PCR identification of *Salmonella* O antigens



+

EIA identification of *Salmonella* H antigens



Molecular serotyping

S. Gaminara
16: d: 1,7

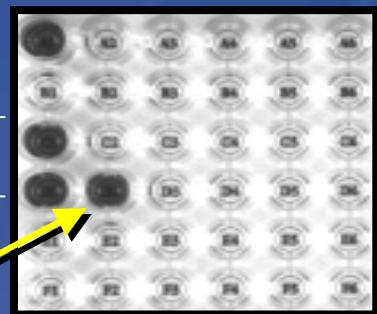
PCR identification of O:16 (I)



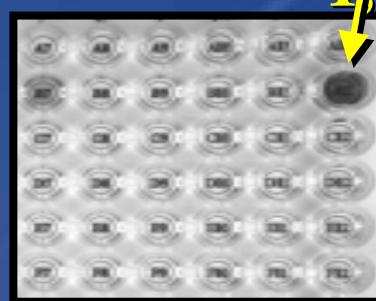
EIA identification of H antigens d and 1,7



Controls



Phase 1



Phase 2

Result: 16: d: 1,7

Conclusions.....so far

- Rapid identification of 15 *Salmonella* serogroups based on PCR amplification of specific gene targets within the *rfb* gene clusters
 - sensitive and specific
 - includes identification of the serotype in ‘rough’ isolates
 - attainable by non-specialized laboratories